

# Variational Method for Estimating the Dipole Moment in the Second Excited State of Fluorescein Molecule from its Electronic UV Absorption Spectra

ALINA COSTINA LUCA<sup>1</sup>, ANA CEZARINA MOROSANU<sup>2</sup>, IRINA MACOVEI<sup>3</sup>, DAN GHEORGHE DIMITRIU<sup>2</sup>,  
DANA ORTANSA DOROHOF\*, IOAN SORIN STRATULAT<sup>4</sup>

<sup>1</sup>Grigore T. Popa University of Medicine and Pharmacy, 16 Universitatii Str., 700115, Iasi, Romania

<sup>2</sup>Alexandru Ioan Cuza University, Faculty of Physics, 11 Carol I Blvd., 700506, Iasi, Romania

<sup>3</sup>Grigore T. Popa University of Medicine and Pharmacy, Faculty of Pharmacy, 16 Universitatii Str., 700115, Iasi, Romania

<sup>4</sup>Grigore T. Popa University of Medicine and Pharmacy, Department of Implantology/Rehabilitation Medicine, Balneology and Physical Therapy, 16 Universitatii Str., 700115, Iasi, Romania

*Electro-optical parameters of fluorescein molecule in the second excited electronic state and information on the interactions with solvents were obtained from a solvatochromic study. Parameters of the solvents such as the refractive index, electrical permittivity and Kamlet-Taft parameters (hydrogen bond acidity and basicity) were related with the experimentally recorded shifts of UV absorption spectral band of fluorescein dissolved in several solvents. Through a variational method, the electric dipole moment and polarizability in excited state of fluorescein molecule were estimated. The calculus requires some parameters of the fluorescein molecule in the ground electronic state, which were determined through a quantum-mechanical study.*

**Keywords:** fluorescein; solvatochromism; dipole moment in the second excited state; UV spectra; quantum-mechanical characterization

Fluorescein is a fluorophore pigment first synthesized by Adolf von Baeyer in 1871 [1] by heating phthalic anhydride and resorcinol over a zinc catalyst. Depending on the pH of the solution, fluorescein exists in seven prototropic forms with different photo-physical properties: three neutral forms and four charged forms [2].

The large rigid  $\pi$ -conjugated structure of fluorescein determines an intense absorption and emission in the visible wavelengths, high photo-stability and high fluorescence quantum yield [3]. These properties make the fluorescein a very good candidate for a wide range of applications. However, it is most widely used as a fluorescence label due to its low cost, high absorptivity and good quantum yield when conjugated to the biomolecules, being used as standard for the calculation of quantum efficiency by relative fluorescence quantum yields methods [4].

Fluorescein is a well-known substance in medicine, being intensively used into a lot of applications. Thus, in ophthalmology, fluorescein is the most used stain, with important applications in both clinical practice and research. Fluorescein is used in investigation of lacrimal characteristics, by observing the normal and pathological pleats of the conjunctiva, and obtaining knowledge about aqueous tear secretion, its complex flow along the lacrimal pool and its clearance [5]. In addition, fluorescein is used in the measurement of intraocular pressure with Goldmann applanation tonometry [6], as well as to observe various ocular surface pathologies. Recently, a new methodology for tear-film dynamics assessment was introduced in ophthalmology and called fluorescein profilometry [7]. Ultra-widefield fluorescein angiography is a technique that enables a visualization of up to 200 degrees of the eye's retina within a single scan [8].

Beside ophthalmology, fluorescein is also widely used in other branches of medicine. Thus, fluorescein fluorescence technology is a valuable tool in the surgical

management of neoplastic and vascular lesions [9]. By using a microscope-integrated fluorescence module, this technique allows performing intraoperative angiography to guide microsurgical management of aneurysms and arteriovenous malformations, as well as the maximization of tumor resection [10]. A comprehensive review on the fluorescein-guided surgery of malignant gliomas can be found in [11]. Dermofluorometry is a technique used for skin microcirculation evaluating, as for example the assessment of diabetic microvascular disease, consisting of measurements of skin fluorescence after an injection of sodium fluorescein [12]. Photobleaching of fluorescein is used as a probe to detect the response of individual cells to oxidative stress and the intracellular reactive oxygen species [13]. As photodynamic sensitizer, fluorescein is of great value in fluorescence immunoassays. Thus, fluorescein derivatives silver nanohybrids have a very good antimicrobial activity against different bacterial strains and fungi [14]. Encapsulated in protective hollow silica nanospheres, fluorescein molecules were used as labels for a biosensor for the rapid detection of *Escherichia coli* O157:H7 [15]. Biocompatible silica-gelatin hybrid aerogels covalently labeled with fluorescein were recently introduced as drug delivery devices [16]. For fluorescent monitoring of drug delivery *in vivo*, fluorescein based chemosensors connected to anticancer drugs were used [17]. Fluorescein is used for synthesization of fluorescein labelled cationic carbosilane dendrimers and dendrons, for biomedical applications (carriers in gene therapy) [18].

Fluorescein is often used in the construction of different kind of sensors for relative humidity, ultraviolet light and temperature monitoring, sensitive blood detection, thiols biological imaging, ammonia and picric acid (2,4,6-trinitrophenol) detection, rapid and selective detection of gold(III),  $\text{Zn}^{2+}$ ,  $\text{Al}^{3+}$ ,  $\text{Cu}^{2+}$ ,  $\text{S}^{2-}$  and  $\text{Fe}^{3+}$  ions, highly selective and sensitive detection of sulfite in aqueous solution and cysteine over other amino acids [19,20].

\* email: ddorohoi@uaic.ro

In optoelectronics, fluorescein dye is well known as a good laser medium in the visible region of the spectrum, being notable for its moderate fluorescence quantum yield and photostability [21]. Nanocrystalline thin films of fluorescein dye were proposed as good candidates for the fabrication of optoelectronic/nonlinear optical devices [22]. Solar cells have been designed based on nanostructured titanium dioxide sensitized with fluorescein sodium salt [23].

Other application of fluorescein dyes are in ground water tracing studies [24], seed security [25], ratiometric fluorescence imaging of nuclear pH in living cells [26] and as an exceptionally simple molecular arithmetic system [27].

Solvatochromism describes the pronounced change in the position and sometimes intensity of a molecular UV-Vis absorption band of solute induced by changes in solvent polarity. Solvatochromic properties of fluorescein have been studied in both aqueous and nonaqueous solvents [3, 28-33]. Based of Lippert's, Bakshiev's and Chamma-Viallet's equations, Acemiođlu *et al.* estimated the excited-state dipole moment in the first excited state of fluorescein in different solvents [31], using information from both absorption and fluorescence spectra. Recently, the dipole moment in the first excited state of fluorescein, as well as the angle between the dipole moments characterizing the electronic states participating to the absorption photons from the visible range were determined [34].

The excited state dipole moment of the molecules can be also estimated based on Abe model [35] but a very great number of parameters even for the solvent and for solute molecule must be known. The electric dipole moments in the electronic states participating to the absorption and emission process were estimated for anthracene using Abe model [36-38].

The aim of this study was to apply the variation method in order to estimate the molecular dipole moment in the second excited electronic state of fluorescein.

## Experimental part

Fluorescein was purchased from Sigma-Aldrich Chemical Company and it was used without purification. The skeletal formula of fluorescein molecule with labeled atoms is displayed in Fig. 1.

Density Functional method of Spartan'14 program [39, 40] was used to compute some theoretical properties of fluorescein. Spartan is a molecular modeling program using

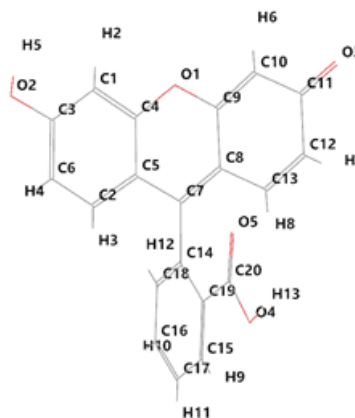


Fig. 1. Skeletal formula of fluorescein molecule with labeled atoms

molecular mechanics and quantum chemical calculations. Spartan offers a variety of graphical tools for interpreting the results of calculations: structure models, molecular orbitals, electron and spin densities, local ionization potentials and electrostatic potentials that can be displayed as surfaces, slices and property maps.

The solvents achieved from Merck were of high spectral

purity. The solvent polarity  $f(n) = \frac{n^2 - 1}{n^2 + 1}$  and polarizability

$$f(\epsilon) = \frac{\epsilon - 1}{\epsilon + 1}$$

functions on refractive index and electric permittivity [41, 42], respectively, are listed in Table 1, which also contains the Kamlet-Taft solvent parameters hydrogen bond donor ( $\alpha$ ) and hydrogen bond acceptor ( $\beta$ ) [43-46].

Ultraviolet electronic absorption spectra of fluorescein were recorded in 14 solvents at room temperature, with QE65000 UV-Vis Ocean-Optics spectrometer.

The statistical analysis of the solvatochromic data was performed by Origin 9 program.

## Results and discussions

The chemical structure of fluorescein was optimized by Spartan'14 program. The lengths of interatomic chemical bonds (calculated with Spartan'14) of fluorescein molecule are listed in Table 2. As it can be seen, the longest chemical bonds are C7-C14 (1.490 Å), C19-C20 (1.489 Å) and C11-C12 (1.471 Å), while the shortest chemical bonds are O2-H5 (0.969 Å) and O5-H1 (0.975 Å).

The measure of some dihedral angles are listed in Table 3. From the data presented here, it results that fluorescein

No.	Solvent	$f(\epsilon)$	$f(n)$	$\beta$	$\alpha$
1	1,4-Dioxane	0.2941	0.2541	0.37	0.00
2	Diethyl ether	0.5261	0.2163	0.47	0.00
3	Chloroform	0.5595	0.2667	0.10	0.20
4	Ethyl acetate	0.6259	0.2273	0.45	0.00
5	Dichloromethane	0.7255	0.2552	0.10	0.13
6	1,2-Dichloroethane	0.7573	0.2661	0.10	0.00
7	1-Hexanol	0.8039	0.2520	0.84	0.80
8	1-Pentanol	0.8113	0.2467	0.86	0.84
9	1-Butanol	0.8462	0.2419	0.84	0.84
10	2-Propanol	0.8631	0.2300	0.84	0.76
11	Ethanol	0.8870	0.2212	0.75	0.86
12	Methanol	0.9135	0.2029	0.66	0.98
13	N,N-Dimethylformamide	0.9225	0.2588	0.69	0.00
14	N,N-Dimethylacetamide	0.9246	0.2625	0.76	0.00

Table 1

SOLVENT POLARITY FUNCTION  $f(\epsilon)$ , SOLVENT POLARIZABILITY FUNCTION  $f(n)$  AND KAMLET-TAFT SOLVENT PARAMETERS ( $\alpha$ - HYDROGEN BOND DONOR,  $\beta$ - HYDROGEN BOND ACCEPTOR)

**Table 2**  
LENGTHS OF INTERATOMIC CHEMICAL BONDS OF FLUORESCIN MOLECULE

Chemical bond	Bond length (Å)	Chemical bond	Bond length (Å)	Chemical bond	Bond length (Å)	Chemical bond	Bond length (Å)
C1-C3	1.390	C2-H3	1.084	C13-C8	1.441	C16-C18	1.390
C3=C6	1.403	C4-O1	1.351	C10-H6	1.083	C18=C14	1.396
C6-C2	1.378	C5-C7	1.438	C11=O3	1.233	C18-H12	1.085
C2=C5	1.407	O1-C9	1.369	C12-H7	1.085	C16-H10	1.085
C5-C4	1.406	C7=C8	1.374	C13-H8	1.085	C17-H11	1.085
C4=C1	1.389	C8-C9	1.446	C7-C14	1.489	C15-H9	1.083
C1-H2	1.085	C9=C10	1.354	C14-C19	1.408	C19-C20	1.484
C3-O2	1.353	C10-C11	1.453	C19=C15	1.399	C20-O4	1.352
O2-H5	0.969	C11-C12	1.471	C15-C17	1.387	C20=O5	1.211
C6-H4	1.083	C12=C13	1.349	C17=C16	1.391	O4-H13	0.974

Dihedral angle	Measure	Dihedral angle	Measure
(C6,C2,C5,C7)	-0.46°	(C5,C7,C14,C19)	-85.29°
(C5,C7,C8,C13)	-179.51°	(C14,C19,C20,O5)	-0.69°
(C10,C11,C12,O3)	179.98°	(C19,C20,O4,H13)	179.75°
(C1,C3,O2,H5)	-0.75°	(C1,C4,O1,C9)	-179.46°

**Table 4**  
MOLECULAR PROPERTIES OF FLUORESCIN, COMPUTED BY SPARTAN'14 PROGRAM

Molecular parameter	Value	Quantitative Structure-Activity Relationship (QSAR)	Value
$E_{\text{HOMO}}$ (eV)	-5.27	Area (Å <sup>2</sup> )	321.44
$E_{\text{LUMO}}$ (eV)	-2.29	Volume (Å <sup>3</sup> )	316.32
Dipole Moment (D)	7.94	PSA (Å <sup>2</sup> )	72.305
Tautomers	3	Log P	-0.91
Conformers	8	Polarizability (Å <sup>3</sup> )	66.33
		HBD Count	2
		HBA Count	4

molecule consists of a plane determined by three cycles connected through a simple covalent bond C7-C14 (Fig. 1) by a benzene cycle substituted at C19 by -COOH atomic group. The benzene cycle is almost perpendicular to the plane determined by the three cycles (the dihedral angle C5,C7,C14,C19 is -85.29°).

The atomic and molecular properties of fluorescein were computed using EDF2/6-31G<sup>\*</sup> density functional and QSAR models, several of them being listed in Table 4.

The charge distribution in a molecule determines its physical and chemical properties. Chemical reactions are associated with charged sites. The most highly-charged site in a molecule is often the most reactive one. Fig. 2 shows the atomic electrostatic charges of fluorescein (expressed in percent of elementary charge), computed by density functional method of Spartan'14.

HOMO (highest occupied molecular orbital) and LUMO (lowest unoccupied molecular orbital) give, by their modulus, the ionization potential and the electron affinity of the fluorescein [47]. They are the most important orbitals with respect to reactivity [47]. The frontier orbitals of fluorescein molecule are shown in Fig. 3a (HOMO surface) and Fig. 3b (LUMO surface), respectively.

**Table 3**  
THE MEASURE OF SOME DIHEDRAL ANGLES OF FLUORESCIN MOLECULE

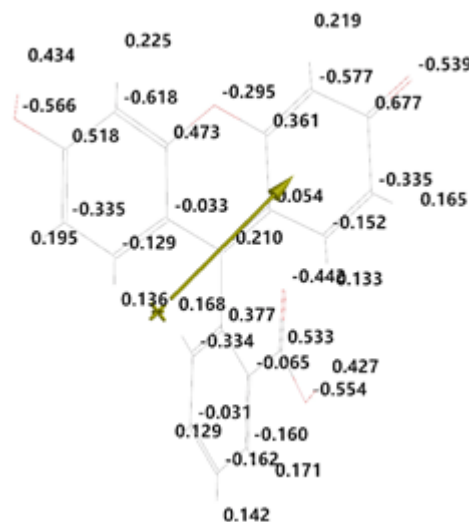


Fig. 2. Electrostatic charges of fluorescein molecule, computed by Spartan program (The arrow indicates the orientation of the dipole moment of the molecule)

The partition coefficient [48] is defined as a particular ratio of the concentrations of a solute between the two solvents. When water is one of the solvents and the other one is a non-polar solvent, then log P value is a measure of hydrophilicity or hydrophobicity. Fluorescein molecule has a hydrophilic character because log P = -0.91 < 0.

The polar surface area (PSA) of a molecule is defined as the surface sum over all polar atoms, primarily oxygen and nitrogen, including also their attached hydrogen atoms. In medical chemistry, PSA is used for the optimization of a drug's ability to permeate cells. Molecules of fluorescein having PSA = 72.3 Å<sup>2</sup> can traverse both the cell membranes (PSA must be less than 140 Å<sup>2</sup>) and the blood-brain barrier (PSA must be less than 90 Å<sup>2</sup>) [49].

Fig. 4 shows the ultraviolet electronic absorption bands of fluorescein in several solvents. The wavenumbers from the maxima of absorption bands were determined for 14 solvents.

The experimental values of the wavenumbers in the maximum of the UV band of fluorescein in different solvents are listed in Tables 5.

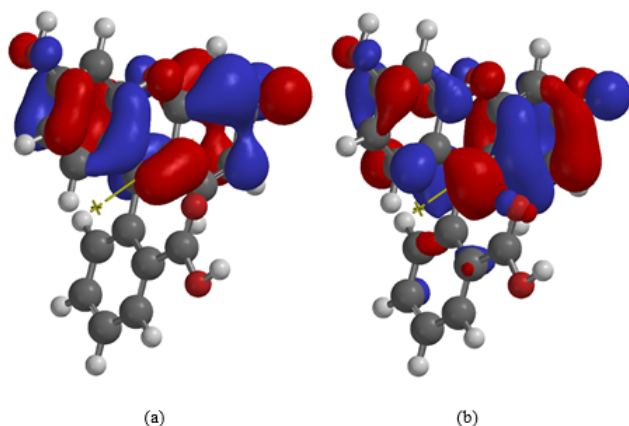


Fig. 3. HOMO (a), LUMO (b) surfaces of fluorescein molecule

**Table 5**  
EXPERIMENTAL AND COMPUTED (USING FORMULA (2)) WAVENUMBERS IN THE MAXIMUM OF UV  
ABSORPTION BAND OF FLUORESCIN

Solvent	$\bar{\nu}_{exp}$ (cm <sup>-1</sup> )	$\bar{\nu}_{calc}$ (cm <sup>-1</sup> )	Solvent	$\bar{\nu}_{exp}$ (cm <sup>-1</sup> )	$\bar{\nu}_{calc}$ (cm <sup>-1</sup> )
1,4-Dioxane	36476.38	36425.90	1-Pentanol	36268.68	36237.29
Diethyl ether	36476.38	36511.64	1-Butanol	36268.68	36252.36
Chloroform	36371.57	36411.50	2-Propanol	36320.05	36294.36
Ethyl acetate	36476.38	36478.76	Ethanol	36320.05	36321.62
DCM	36476.38	36442.62	Methanol	36371.57	36373.17
1,2-DCE	36423.24	36411.50	N,N-DMF	36371.57	36329.89
1-Hexanol	36165.06	36231.36	N,N-DMA	36268.68	36306.85

The total shift of the electronic bands can be described by the following relation [50-53]:

$$\bar{\nu}_{calc} = \bar{\nu}_0 + C_1 f(\epsilon) + C_2 f(n) + C_3 \beta + C_4 \alpha, \quad (1)$$

where  $\bar{\nu}_0$  (cm<sup>-1</sup>) has the significance of the wavenumber in the maximum of absorption electronic band for gaseous phase of the spectrally active substance and  $C_1$ - $C_4$  are the regression coefficients obtained by multilinear regression analysis. The terms  $C_1 f(\epsilon)$  and  $C_2 f(n)$  give the contribution of the universal interactions by orientation-induction and polarization-dispersion processes, respectively, while the terms  $C_3 \beta$  and  $C_4 \alpha$  give the contribution of the specific interactions to the total spectral shift.

When the solvent parameters and the experimental wavenumber of the absorption band are known, a multilinear regression applied to formula (1) allow estimating the regression coefficients from Table 6.

Using the values of these coefficients, the relation (1) can be written as:

$$\bar{\nu}_{calc} (\text{cm}^{-1}) = 37232.1 - 57.5 f(\epsilon) - 2797.2 f(n) - 181.2 \beta - 121.6 \alpha, \quad (2)$$

The computed (by using the relation (2)) wavenumbers in the maxima of UV band are listed in Table 5. The computed wavenumbers  $\bar{\nu}_{calc}$  (cm<sup>-1</sup>) versus the experimental wavenumbers  $\bar{\nu}_{exp}$  (cm<sup>-1</sup>) are plotted in Fig. 5. A good linear dependence between the experimental and calculated values of the wave numbers was obtained: the slope is 0.88, the regression coefficient is  $R = 0.87$  and the standard deviation is  $SD = 33.19$ .

The contribution of each type of interaction to the total spectral shift in different solvents, in cm<sup>-1</sup> and in percentage (%), is listed in Table 7. The data show that the dispersion-polarization interactions have the greatest contribution to

**Table 6**  
REGRESSION COEFFICIENTS OBTAINED BY MULTILINEAR REGRESSION ANALYSIS FOR ELECTRONIC ABSORPTION BAND  
OF FLUORESCIN IN ULTRAVIOLET

Intercept		$C_1$ (cm <sup>-1</sup> )		$C_2$ (cm <sup>-1</sup> )		$C_3$ (cm <sup>-1</sup> )		$C_4$ (cm <sup>-1</sup> )		Statistics	
Value	Error	Value	Error	Value	Error	Value	Error	Value	Error	Adj. R-Square	Root-MSE (SD)
37232.07	164.44	-57.55	78.44	-2797.18	651.01	-181.22	55.38	-121.64	39.04	0.826	40.86



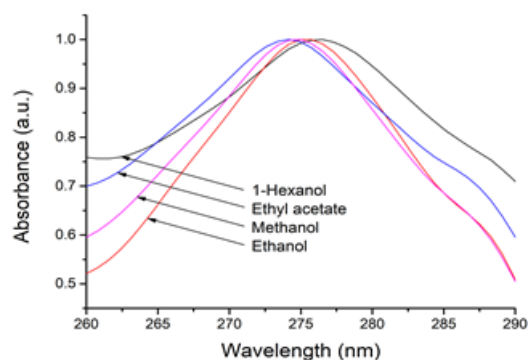


Fig. 4. Ultraviolet electronic absorption band of fluorescein in four solvents (1-hexanol, ethyl acetate, methanol and ethanol, respectively)

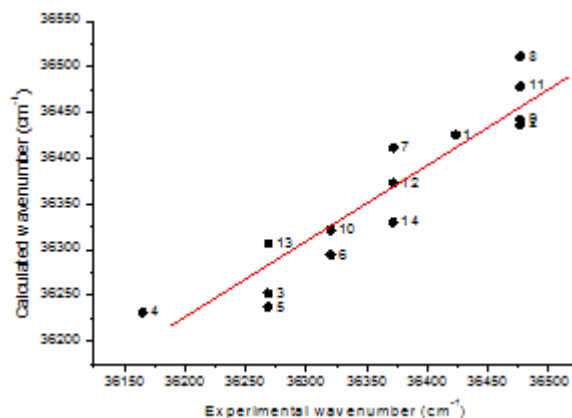


Fig. 5.  $\bar{\nu}_{\text{calc}}$  (calculated using relation (2)) vs.  $\bar{\nu}_{\text{exp}}$  for the UV electronic absorption band of fluorescein

Solvent	$C_1 f(\epsilon)$ (cm <sup>-1</sup> )	$p_1$ %	$C_2 f(n)$ (cm <sup>-1</sup> )	$p_2$ %	$C_3 \beta$ (cm <sup>-1</sup> )	$p_3$ %	$C_4 \alpha$ (cm <sup>-1</sup> )	$p_4$ %
1,4-Dioxane	16.9	2.1	710.8	89.4	67.1	8.4	0.0	0.0
Diethyl ether	30.3	4.2	605.0	84.0	85.2	11.8	0.0	0.0
Chloroform	32.2	3.9	745.9	90.9	18.1	2.2	24.3	3.0
Ethyl acetate	36.0	4.8	635.7	84.4	81.5	10.8	0.0	0.0
DCM	41.8	5.3	713.8	90.4	18.1	2.3	15.8	2.0
1,2-DCE	43.6	5.4	744.5	92.3	18.1	2.2	0.0	0.0
1-Hexanol	46.3	4.6	704.9	70.4	152.2	15.2	97.3	9.7
1-Pentanol	46.7	4.7	690.1	69.4	155.8	15.7	102.2	10.3
1-Butanol	48.7	5.0	676.6	69.1	152.2	15.5	102.2	10.4
2-Propanol	49.7	5.3	643.4	68.6	152.2	16.2	92.4	9.9
Ethanol	51.0	5.6	618.9	68.0	135.9	14.9	104.6	11.5
Methanol	52.6	6.1	567.5	66.1	119.6	13.9	119.2	13.9
N,N - DMF	53.1	5.9	724.0	80.3	125.0	13.9	0.0	0.0
N,N - DMA	53.2	5.8	734.3	79.4	137.7	14.9	0.0	0.0

**Table 7**  
CONTRIBUTION OF EACH  
TYPE OF INTERACTION TO  
THE TOTAL SPECTRAL SHIFT  
IN ULTRAVIOLET DOMAIN

the total spectral shift. The weight of dispersion-polarization interactions is between 66.1 and 92.3 %.

If only the electronic absorption spectra of a molecule in different solvents are available, a variational method could be applied to obtain information on both the electric dipole moment in the excited state of the studied molecule and the angle between the dipole moments in the electronic states participating in absorption process. Such a procedure, described below, was successfully applied for other molecules [52-56].

The regression coefficients  $C_1$  and  $C_2$  from relation (1) depend on the microscopic parameters of the solute molecule [52, 53]:

$$C_1 = \frac{2\mu_g(\mu_g - \mu_e \cos \varphi)}{hca^3} + 3kT \frac{\alpha_g - \alpha_e}{a^3} \quad (4)$$

$$C_2 = \frac{\mu_g^2 - \mu_e^2}{hca^3} - \frac{2\mu_g(\mu_g - \mu_e \cos \varphi)}{hca^3} - 3kT \frac{\alpha_g - \alpha_e}{a^3} + \frac{3}{2} \frac{\alpha_g - \alpha_e}{a^3} \frac{I_u I_v}{I_u + I_v}$$

Using Eqs. (3) and (4), the dipole moment and the polarizability of the spectrally active molecule in excited state can be estimated. Thus, from Eqs. (3) and (4) the next relation can be obtained:

$$C_1 + C_2 = \frac{\mu_g^2 - \mu_e^2}{hca^3} + \frac{3}{2} \frac{\alpha_g - \alpha_e}{a^3} \frac{I_u I_v}{I_u + I_v} \quad (5)$$

The molecular radius  $a$  can be calculated function of the values of volume and surface area in the ground state of the molecule (Table 1):

$$a = \frac{3V}{A} \quad (6)$$

For the fluorescein molecule, the radius can be approximated by  $a = 2.9522 \text{ \AA} = 2.9522 \cdot 10^{-8} \text{ cm}$ .

In Eqs. (3)-(5) the following notations were made:  $\mu_g$  and  $\mu_e$  - the dipole moments of the spectrally active molecule in the ground and excited states, respectively,  $\varphi$  - the angle between them,  $\alpha_g$  and  $\alpha_e$  - the polarizabilities of the molecule in its ground and excited states, respectively,  $c$  - the speed of light in vacuum,  $a$  - the radius of molecule,  $h$  - the Planck's constant,  $k$  - the Boltzmann's constant,  $T$  - the absolute temperature,  $I_u$  - the ionization potential of the spectrally active molecule,  $I_v$  - the ionization potential of the solvent molecule. Chloroform ( $I_v = 8.4 \text{ eV}$ ) was the solvent used to calculate the polarizability and the dipole moment of fluorescein molecule in the excited state.

The following data are used in the calculus:  $\mu_g = 7.94 \text{ D}$ ,  $\alpha_g = 66.33 \text{ \AA}^3$ ,  $a = 2.9522 \text{ \AA}$ ,  $I_u = -E_{\text{HOMO}} = 5.27 \text{ eV}$ ;  $I_v = 8.4 \text{ eV}$ ;  $C_1 = -57.55 \text{ cm}^{-1}$ ;  $C_2 = -2797.18 \text{ cm}^{-1}$  (Tables 1 and 6). One obtains the following equations:

Angle $\varphi$ (degrees)	$\mu_e$ (D)	$\alpha_e$ (Å <sup>3</sup> )	Angle $\varphi$ (degrees)	$\mu_e$ (D)	$\alpha_e$ (Å <sup>3</sup> )
0	7.94	68.52	30	9.20	65.28
10	8.06	68.23	40	11.78	57.11
20	8.46	67.24	50	12.52	54.39
25.3	8.73	66.54	60	16.36	37.72
25.4	8.81	66.33	70	25.14	17.20
25.5	8.82	66.31			

**Table 8**  
EXCITED STATE DIPOLE MOMENT AND  
POLARIZABILITY FOR DIFFERENT ANGLES  
 $\varphi \in (0, 79^\circ)$  FOR UV ABSORPTION BAND OF  
FLUORESCENIN

$$\alpha_e = 78.0297 - 0.1507\mu_e^2, \quad (7)$$

$$0.0183\mu_e^2 - 15.88\mu_e \cos\varphi + 124.9647 = 0. \quad (8)$$

In Eqs. (7) and (8)  $\alpha_e$  is expressed in Å<sup>3</sup>, while  $\mu_e$  is expressed in D. Equation (8) has real solutions for positive discriminator  $\Delta$ :

$$\Delta = 252.1744\cos^2\varphi - 9.1474. \quad (9)$$

The next condition is necessary to have real values for excited state dipole moment:

$$\Delta \geq 0 \text{ for } \varphi \leq 79.02^\circ. \quad (10)$$

It results that the angle  $\varphi$  between the molecular dipole moments in the electronic states of fluorescein must be smaller than 79°. Considering the value 7.94 D for the ground state dipole moment and 66.33 Å<sup>3</sup> for the ground state polarizability of fluorescein, one obtains the data from Table 8 for step by step variation of  $\varphi$  with 10°.

For  $\varphi \geq 25.5^\circ$  the molecular polarizability is smaller than the ground state polarizability of fluorescein. The electronic transition in UV absorption takes place at this angle.

## Conclusions

Fluorescein molecule has been investigated by quantum mechanical and spectral measurements in order to determine its excited state dipole moment using only the information from electronic absorption spectrum. In UV domain of the fluorescein absorption spectrum, a  $\pi \rightarrow \pi^*$  transition takes place, with increase of the molecular dipole moment.

Based on the quantum mechanical and spectral results, the dipole moment of fluorescein in the second excited electronic state  $\pi^*$  is determined in this paper in the hypothesis that the molecular electric polarizability does not change its value in the absorption process. The angle between the dipole moments in the ground and excited states of fluorescein was also estimated.

## References

- VON BAEYER, A., Berichte der Deutschen Chemischen Gesellschaft zu Berlin, **4**, 1871, p. 555.
- MCQUEEN, P. D., SAGOO, S., YAO, H., JOCKUSCH, R. A., Angew. Chem., **122**, 2010, p. 9379.
- NADERI, F., FARAJTABAR, A., J. Mol. Liq., **221**, 2016, p. 102.
- BROUWER, A. M., Pure Appl. Chem., **83**, 2011, p. 2213.
- MURUBE, J., Ocul. Surf., **11**, 2013, p. 212.
- MURUBE, J., Ocul. Surf., **11**, 2013, p. 144.
- GARASZCZUK, I. K., ISKANDER, D. R., Cont. Lens Anterior Eye, **40**, 2017, p. 208.
- KASHANI, A. H., CHEN, C.-L., GAHM, J. K., ZHENG, F., RICHTER, G. M., ROSENFELD, P. J., SHI, Y., WANG, R. K., Prog. Retin. Eye Res., **60**, 2017, p. 66.
- REY-DIOS, R., COHEN-GADOL, A. A., Acta Neurochir., **155**, 2013, p. 701.

- KAKUCS, C., FLORIAN, I.-A., UNGUREANU, G., FLORIAN, I.-S., World Neurosurg., **105**, 2017, p. 406.
- ACERBI, F., CAVALLO, C., BROGGI, M., CORDELLA, R., ANGHILERI, E., EOLI, M., SCHIARITI, M., BROGGI, G., FERROLI, P., Neurosurg. Rev., **37**, 2014, p. 547.
- OH, D. K., JONES, R. N., MARSHALL, W., MAGIN, R. L., Skin Res. Technol., **7**, 2001, p. 105.
- PLATKOV, M., TIROSH, R., KAUFMAN, M., ZURGIL, N., DEUTSCH, M., J. Photochem. Photobiol. B-Biol., **140**, 2014, p. 306.
- NEGM, N. A., ABOU KANA, M. T. H., ABD-ELAAL, A. A., ELWAHY, A. H. M., J. Photochem. Photobiol. B-Biol., **162**, 2016, p. 421.
- HU, R.-R., YIN, Z.-Z., ZENG, Y.-B., ZHANG, J., LIU, H.-Q., SHAO, Y., REN, S.-B., LI, L., Biosens. Bioelectron., **78**, 2016, p. 31.
- VERES, P., KIRALI, G., NAGY, G., LAZAR, I., FABIÁN, I., KALMAR, J., J. Non-Cryst. Solids, **473**, 2017, p. 17.
- BAZYLEVICH, A., PATSENKER, L. D., GELLERMAN, G., Dyes Pigments, **139**, 2017, p. 460.
- FUENTES-PANIAGUA, E., SERRAMIA, M. J., SANCHEZ-NIEVES, J., ÁLVAREZ, S., MUNOZ-FERNANDEZ, M. A., GOMEZ, R., DE LA MATA, F. J., Eur. Polym. J., **71**, 2015, p. 61.
- BHATTACHARYA, S., AGARWAL, A. K., CHANDA, N., PANDEY, A., SEN, A. K. (Eds.), Environmental, Chemical and Medical Sensors, Springer Nature, Singapore, 2018.
- WEBSTER, J. G., EREN, H. (Eds.), Measurement, Instrumentation, and Sensors Handbook. Electromagnetic, Optical, Radiation, Chemical, and Biomedical Measurement, CRC Press, Taylor & Francis Group, Boca Raton, FL, 2014.
- AL-AQMAR, D. M., ABDELKADER, H. I., ABOU KANA, M. T. H., Opt. Mater., **47**, 2015, p. 573.
- ABUTALIB, M. M., SHKIR, M., YAHIA, I. S., ALFAIFY, S., EL-NAGGAR, A. M., GANESH, V., Optik, **127**, 2016, p. 6601.
- FAROOQ, W. A., FATEHMULLA, A., YAKUPHANOGU, F., YAHIA, I. S., ALI, S. M., ATIF, M., ASLAM, M., TAWFIK, W., Theor. Exp. Chem., **50**, 2014, p. 121.
- CHUA, L. H. C., ROBERTSON, A. P., YEE, W. K., SHUY, E. B., LO, E. Y. M., LIM, T. T., TAN, S. K., Ground Water, **45**, 2007, p. 85.
- GUAN, Y. J., WANG, J. C., HU, J., LI, Y. P., MA, W. G., HU, W. M., ZHU, S. J., Ind. Crops Prod., **45**, 2013, p. 367.
- NAKAMURA, A., TSUKIJI, S., Bioorganic Med. Chem. Lett., **27**, 2017, p. 3127.
- MARGULIES, D., MELMAN, G., SHANZER, A., Nat. Mater., **4**, 2005, p. 768.
- CHOI, M. F., HAWKINS, P., Spectrosc. Lett., **27**, 1994, p. 1049.
- KLONIS, N., CLAYTON, A. H. A., VOSS JR., E. W., SAWYER, W. H., Photochem. Photobiol., **67**, 1998, p. 500.
- BISWAS, S., BHATTACHARYA, S. C., SEN, P. K., MOULIK, S. P., J. Photochem. Photobiol. A-Chem., **123**, 1999, p. 121.
- ACEMIÖDLÜ, B., ARIK, M., EFEOÖDLÜ, H., ONGANER, Y., J. Mol. Struct.-Theochem, **548**, 2001, p. 165.
- MCHEDLOV-PETROSSYAN, N. O., IVANOV, V. V., Russ. J. Phys. Chem. A, **81**, 2007, p. 112.
- NADERI, F., FARAJTABAR, A., GHARIB, F., J. Mol. Liq., **190**, 2014, p. 126.
- MOROSANU, A. C., DIMITRIU, D. G., DOROHAI, D. O., J. Mol. Struct., **1180**, 2019, p. 723.

- 35.ABE, T., Bull. Chem. Soc. Japan, **38**, 1965, p. 1314 and **39**, 1966, p. 936.
- 36.DOROHAI, D. O., DIMITRIU, D. G., Studia Universitatis Babes Bolyai, Physica, Special Issue PIM 2001, 2001, p. 337.
- 37.TIGOIANU, R.I., DOROHAI, D.O., AIRINEI, A., Rev. Chim. (Bucharest), **60**, no. 1, 2009, p. 42.
- 38.TIGOIANU, R.I., AIRINEI, A., DOROHAI, D.O., Rev. Chim. (Bucharest), **60**, no. 5, 2010, p. 491.
- 39.YOUNG, D., Computational Chemistry: A Practical Guide for Applying Techniques to Real World Problems, John Wiley & Sons, New York, 2001.
- 40.HEHRE, W., OHLINGER, S., Spartan'14 for Windows, Macintosh and Linux, Tutorial and User's Guide, Wavefunction Inc., Irvine, CA, 2014.
- 41.BAKHSHIEV, H. G., Spectroscopy of Intermolecular Interactions (in Russian), Nauka, Leningrad, 1972.
- 42.HURJUI, I., IVAN, L. M., DOROHAI, D. O., Spectrochim. Acta A, **102**, 2013, p. 219.
- 43.\*\*\* <http://www.stenutz.eu/chem/solv26.php> (accessed on December 17, 2018).
- 44.GHEORGHIES, L. V., DIMITRIU, M., FILIP, E., DOROHAI, D. O., Rom. J. Phys., **55**, 2010, p. 103.
- 45.GHEORGHIES, C., GHEORGHIES, L. V., DOROHAI, D. O., J. Mol. Struct., **887**, 2008, p. 122.
- 46.BABUSCA, D., DOROHAI, D. O., Spectrochim. Acta A, **152**, 2016, p. 149.
- 47.FLEMING, I., Frontier orbitals and organic chemical reactions, John Wiley & Sons, Chichester, 1976.
- 48.PARTHASARATHI, R., SUBRAMANIAN, V., ROY, D. R., CHATTARAJ, P. K., Bioorgan. Med. Chem., **12**, 2004, p. 5533.
- 49.HITCHCOCK, S. A., PENNINGTON, L. D., J. Med. Chem., **49**, 2006, p. 7559.
- 50.DOROHAI, D. O., DIMITRIU, D. G., DIMITRIU, M., CLOSCA, V., J. Mol. Struct., **1044**, 2013, p. 79.
- 51.DOROHAI, D. O., Ukr. J. Phys., **63**, 2018, p. 701.
- 52.BABUSCA, D., MOROSANU, A. C., BENCHEA, A. C., DIMITRIU, D. G., DOROHAI, D. O., J. Mol. Liq., **269**, 2018, p. 940.
- 53.MOROSANU, A. C., GRITCO-TODERASCU, A., CREANGA, D. E., DOROHAI, D. O., Spectrochim. Acta A, **189**, 2018, p. 307.
- 54.BABUSCA, D., DIMITRIU, D. G., DOROHAI, D. O., Anal. Lett., **49**, 2016, p. 2615.
- 55.BABUSCA, D., BENCHEA, A. C., DIMITRIU, D. G., DOROHAI, D. O., Anal. Lett., **50**, 2017, p. 2740.
- 56.MOROSANU, A. C., BENCHEA, A. C., BABUSCA, D., DIMITRIU, D. G., DOROHAI, D. O., Anal. Lett., **50**, 2017, p. 2725.

---

Manuscript received: 15.01.2019